

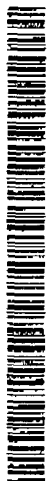
(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau(43) International Publication Date
25 May 2001 (25.05.2001)

PCT

(10) International Publication Number
WO 01/36979 A2

- (51) International Patent Classification: G01N 33/68 (74) Agents: CORNISH, K., V., J. et al.; Kilburn & Stoebe, 20 Red Lion Street, London WC1R 4PJ (GB).
- (21) International Application Number: PCT/GB00/04315
- (22) International Filing Date: 10 November 2000 (10.11.2000)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data: 9927125.6 16 November 1999 (16.11.1999) GB
- (71) Applicant (for all designated States except US): UNIVERSITY OF READING [GB/GB]; Whiteknights House, Whiteknights, Reading RG6 6AH (GB).
- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, GR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).
- Published:
— Without international search report and to be republished upon receipt of that report.
- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 01/36979 A2

(54) Title: PLACENTAL HUMAN NEUROKININ B PRECURSOR

(57) Abstract: Methods of diagnosing pregnancy induced hypertension or pre-eclampsia by the measurement of the production of neurokinin B, its precursor and fragments thereof are provided, as are kits for use in the methods. Treatments of the conditions and methods of preparing suitable medicaments are also provided as are antibodies and useful antigenic materials.

WO 01/36979

PCT/GB00/04315

1

PLACENTAL HUMAN NEUROKININ B PRECURSOR

The present invention is concerned with the detection of the production of the human precursor of neurokinin B by the placenta and to the detection of the
5 production of neurokinin B gene products, or variants, or fragments thereof as a means of predicting the onset of pregnancy induced hypertension or pre-eclampsia or related foetal complications (or following their course). The application is also directed to methods of preventing or treating pregnancy-induced hypertension or pre-eclampsia by suppressing the effects of
10 excessive neurokinin B secreted into maternal blood.

Pregnancy-induced hypertension (PIH) and pre-eclampsia, two of the most elusive and complex conditions of pregnancy, have been very difficult to define and manage. Pre-eclampsia is still one of the most common and life
15 threatening complications of pregnancy in the Western World. The primary cause of pre-eclampsia has been difficult to elucidate because its signs and symptoms have always presented as a cluster of conditions. Hence, it has been defined as a syndrome, commonly presenting with the features of maternal hypertension and proteinuria, but including extensive complications
20 involving the maternal liver, coagulation and nervous systems (Henriksen, T., (1998) Scand. J. Rheumatol. Suppl. 107 86-91). The clinical problems of pre-eclampsia normally become apparent only in the second half of pregnancy and are believed to emerge during the first trimester. It would appear that pre-eclamptic complications only present if placental tissue is
25 present in the uterus of the mother. Indeed, cases of hydatidiform mole can present with pre-eclampsia where the uterus only contains disordered placental tissue (Nugent, C.E. et al (1966) Obstet. Gynecol. 87 829-31). Once pre-eclampsia is diagnosed during the course of pregnancy and the placental tissue is surgically removed or expelled during birth the condition
30 ultimately clears. There have been many suggestions about the causes of pre-eclampsia ranging from the development of a poor placental/uterine vascular system to the immunology of incompatibility between the mother

WO 01/36979

PCT/GB00/04315

2

and foetus. Though these theories do have some substance they do not account for the systemic effects of this syndrome. Many symptoms are likely to be the result of secondary effects of hypertension and not the direct cause of the syndrome. Early detection of the development of PIH or pre-eclampsia would therefore be of great benefit in allowing precautionary measures to be taken, including specific treatment of hypertension and other complications associated with pre-eclampsia such as seizures, blot clotting problems etc.

The placental damage visible and hypertension observed in an expectant mother with pre-eclampsia has been implicated in an increased risk of foetal complications including growth retardation and foetal hypoxia. In extreme cases this could be a cause of miscarriage. In other studies, pre-eclampsia has been postulated as a maternal and foetal adaptation to foetal growth retardation. Since not all women with foetal growth retardation develop pre-eclampsia the decisive factor is a maternal response (Walker, J. (2000) The Lancet 356 1260-1265). Characteristics of this adaptation are present in not only pre-eclampsia but also in foetal growth retardation and miscarriage. For example, the failure of the normal expansion of plasma volume in the mother is associated with both impaired foetal growth and pre-eclampsia (Gulmezoglu AM, Hofmeyr GJ (2000) Cochrane Database Syst Rev 2 CD000167). Problems observed in pre-eclampsia such as thrombophilia are suggested to be the result of thrombotic lesions in a pathological placenta (Mousa HA, Alfrevici Z (2000) Hum Reprod 15:1830-3). It is apparent therefore that pre-eclampsia and foetal growth retardation and foetal hypoxia are linked, and diagnostic methods and treatments for pre-eclampsia may also be suitable in the prediction, diagnosis and/or treatment of these foetal conditions

Neurokinin B (NKB) belongs to a family of peptides called tachykinins, the first and most well known of which is substance P which was discovered in 1931 (von Euler, U.S. and Gaddum, J.H. (1931) J Physiol 72:74-87). It took over another five decades before the discovery of a further two members of

WO 01/36979

PCT/GB00/04315

3

the tachykinin family, one designated substance K or neurokinin A (Kimura, S., *et al* (1983) Proc. Japan Acad 59B 101-104) and the other designated neuromedin K, now know as neurokinin B (Kangawa, K., *et al* (1983). Biochem. Biophys. Res. Commun. 114 533-540). The tachykinins have been

5 implicated to have a wide variety of biological actions from smooth muscle contraction, vasodilation, pain transmission, neurogenic inflammation, to the activation of the immune system (Longmore, J., *et al* (1997) Canadian J. Physio & Pharmacol. 75 612-621). Neurokinin B has been found to be the most potent neurokinin to cause vasoconstriction of both the mesentenc

10 vascular bed (D'Orleans-Juste, P. *et al* (1991). Eur. J. Pharmacol. 204 329-334) and contraction of the hepatic portal vein (Mastrangelo, D., *et al* (1987) Eur J Pharmacol. 134, 321-6). Neurokinin B is also the most potent member of the family to act at the NK₃ receptor and, whilst substance P and K slow down the heart rate, NK₃ receptor agonists have the opposite effect in that

15 they increase heart rate when perfused in the canine coronary arterial blood supply (Thompson, G.W. *et al* (1998) American Journal of Physiology-Regulatory Integrative and Comparative Physiology 275 (5), 1683-1689). In an animal model, intravenous injections of neurokinin B in guinea pigs have been shown to produce a dose related hypertension, and very high levels of

20 neurokinin B agonist led to animal discomfort (Roccon, A., *et al* (1996) Brit J Pharmacol. 118 1095-1102). Similar experiments have shown an increase in blood pressure upon intravenous infusion of neurokinin B in rats (Page *et al.*, (2000) Nature 405 797-800). Neurokinin B has not been reliably found in any peripheral tissues taken from experimental animals; for example, Moussaoui

25 *et al* (Neuroscience (1992) 48, 967-978) tested a wide range of peripheral tissues using a very sensitive and specific assay system and found no trace of neurokinin B at all.

A human neurokinin B precursor has been identified which, on processing,

30 gives rise to a peptide identical to neurokinin B of other mammalian species (bovine, porcine, rat and mouse) (Incyte Pharmaceuticals Inc., international patent application no WO98/57986). We have discovered, most surprisingly,

WO 01/36979

PCT/G800/04315

4

that this human neurokinin B precursor is produced by placental tissue during pregnancy and that neurokinin B and fragments of the precursor are passed into the maternal bloodstream.

- 5 We have found that in normal pregnancy, substantial levels (eg 100 picomolar range) of neurokinin B (and other breakdown products of the human neurokinin B precursor) are found in the maternal blood stream near to term, but that zero or very low levels are found before this. However, in some cases near term levels are identified at an early stage of pregnancy (eg
10 after only 9 weeks), and in cases of pregnancy induced hypertension or pre-eclampsia very high (nanomolar) concentrations of neurokinin B are found in the maternal plasma near to term. Thus, detection of raised plasma levels of neurokinin B, neurokinin B precursor, its breakdown products, or variants thereof at an early stage will provide an indication of the likely development
15 of pregnancy induced hypertension or pre-eclampsia and may even provide an indication of the likely future severity of these conditions. Furthermore, reduction in the levels of circulating neurokinin B (or reduction of its effects) will ameliorate the adverse effects upon the mother seen in these conditions. As a result of the relationship between pre-eclampsia and foetal
20 complications including foetal growth retardation and/or foetal hypoxia, neurokinin B agonists or antagonists may be useful in ameliorating these conditions. Overproduction of the human neurokinin B precursor may also be a causative factor in certain hypertensive conditions in non-pregnant individuals (either through the effect of neurokinin B or one or more of the
25 other breakdown products of the precursor).

- In a first aspect of the invention there is provided a method of predicting pregnancy induced hypertension in a human subject by assessing the concentration in a biological sample, e.g. blood, of a human neurokinin B
30 precursor gene product or a variant or a fragment thereof.

WO 01/36979

PCT/CB00/04315

5

In a second aspect of the invention there is provided a method of predicting pre-eclampsia or related foetal complications in a human subject by assessing the concentration in a biological sample, e.g. blood, of a human neurokinin B precursor gene product or a variant or a fragment thereof.

5

In a third aspect of the invention there is provided a method of diagnosing pregnancy induced hypertension in a human subject by assessing the concentration in a biological sample, e.g. blood, of a human neurokinin B precursor gene product or a variant or a fragment thereof.

10

In a fourth aspect of the invention there is provided a method of diagnosing pre-eclampsia or related foetal complications in a human subject by assessing the concentration in a biological sample, e.g. blood, of a human neurokinin B precursor gene product or a variant or a fragment thereof.

15

Preferably, the methods of the first, second, third or fourth aspects comprise assessing the concentration in a biological sample, e.g. blood, of neurokinin B.

20

In a fifth aspect of the invention there is provided a method of estimating the likely future degree of pregnancy induced hypertension in a human subject by assessing the concentration in a biological sample, e.g. blood, of human neurokinin B precursor gene product or a variant or a fragment thereof, and correlating the result with the predicted future severity of pregnancy induced

25

hypertension.

30

In a sixth aspect of the invention there is provided a method of estimating the likely future degree of pre-eclampsia or related foetal complications in a human subject by assessing the concentration in a biological sample, e.g. blood, of human neurokinin B precursor or a variant or a fragment thereof, and correlating the result with the predicted future severity of pre-eclampsia or related foetal complications

WO 01/36979

PCT/GB00/04315

6

Preferably, the methods of the fifth and sixth aspects comprise assessing the concentration in a biological sample, e.g. blood, of neurokinin B, and correlating the result with the predicted future severity of pregnancy induced
5 hypertension or pre-eclampsia or related foetal complications, respectively.

In a seventh aspect of the invention there is provided a method of preventing or treating pregnancy induced hypertension in a human subject by the administration of an agent which inhibits the biological effect of neurokinin B
10

in an eighth aspect of the invention there is provided a method of preventing or treating pre-eclampsia or related foetal complications in a human subject by the administration of an agent which inhibits the biological effect of neurokinin B
15

In a ninth aspect of the invention there is provided the use of a human neurokinin B precursor gene product or a variant or a fragment thereof in the manufacture of a diagnostic for use in the prediction or diagnosis of pregnancy-induced hypertension
20

In a tenth aspect of the invention there is provided the use of a human neurokinin B precursor gene product or a variant or a fragment thereof in the manufacture of a diagnostic for use in the prediction or diagnosis of pre-eclampsia or related foetal complications.
25

Preferably, the ninth and tenth aspects comprise the use of an epitopic variant or epitopic fragment of human neurokinin B precursor. More preferably, the methods comprise the use of neurokinin B in the manufacture of a diagnostic for use in the prediction or diagnosis of pregnancy induced
30 hypertension, pre-eclampsia or related foetal complications.

WO 01/36979

PCT/GB00/04315

7

In an eleventh aspect of the invention there is provided the use of an agent which inhibits the biological effect of neurokinin B in the manufacture of a medicament for the prevention or treatment of pregnancy induced hypertension.

5

In a twelfth aspect of the invention there is provided the use of an agent which inhibits the biological effect of neurokinin B in the manufacture of a medicament for the prevention or treatment of pre-eclampsia or related foetal complications.

10

In a preferred embodiment of the eleventh and twelfth aspects, there is provided a pharmaceutical composition comprising an agent which inhibits the biological effect of neurokinin B, for use in the prevention or treatment of pregnancy induced hypertension, pre-eclampsia or related foetal complications.

15

In a thirteenth aspect of the invention there is provided a kit for the prediction or diagnosis of pregnancy induced hypertension comprising a binding partner, eg an antibody, to a neurokinin B precursor gene product or variant or fragment thereof.

20

In a fourteenth aspect of the invention there is provided a kit for the prediction or diagnosis of pre-eclampsia or related foetal complications comprising a binding partner, eg an antibody, to a neurokinin B precursor gene product or variant or fragment thereof.

25

In a fifteenth aspect of the invention there is provided a kit for the prediction or diagnosis of pregnancy induced hypertension, comprising a binding partner, eg an antibody, to a neurokinin B precursor gene product or variant or fragment thereof, together with instructions for the performance of an assay for predicting the levels of neurokinin B in a biological sample and

30

WO 01/36979

PCT/GB00/04315

8

correlating the assay results with the likely future development of pregnancy induced hypertension.

In a sixteenth aspect of the invention there is provided a kit for the prediction
5 or diagnosis of pre-eclampsia or related foetal complications, comprising a
binding partner, eg an antibody, to neurokinin B precursor gene product or
variant or fragment thereof, together with instructions for the performance of
an assay for predicting the levels of neurokinin B in a biological sample and
correlating the assay results with the likely future development of pre-
10 eclampsia or related foetal complications.

In a seventeenth aspect of the invention there is provided a kit for use in
estimating the likely future degree of pregnancy induced hypertension,
comprising a binding partner, eg an antibody, to a neurokinin B precursor
15 gene product or variant or fragment thereof, together with instructions for the
performance of an assay for predicting the levels of neurokinin B in a
biological sample and correlating the assay results with the predicted future
severity of pregnancy induced hypertension.

20 In an eighteenth aspect of the invention there is provided a kit for use in
estimating the likely future degree of pre-eclampsia or related foetal
complications, comprising a binding partner, eg an antibody, to a neurokinin
B precursor gene product or variant or fragment thereof, together with
instructions for the performance of an assay for predicting the levels of
25 neurokinin B in a biological sample and correlating the assay results with the
predicted future severity of pre-eclampsia or related foetal complications.

Preferably, the kits of the thirteenth to eighteenth aspects of the invention
comprise a binding partner, e.g. an antibody, to a neurokinin B precursor,
30 neurokinin B or epitopic variants or epitopic fragments thereof. More
preferably the kits comprise a binding partner to the polypeptide sequences
of Figures 1 or 2, or epitopic variants or epitopic fragments thereof.

WO 01/36979

PCT/GB00/04315

9

In a nineteenth aspect of the invention there is provided the use of an agonist of neurokinin B or neurokinin B in the preparation of a medicament for the reduction of blood volume in cases of hypotension.

5

In a twentieth aspect of the invention there is provided the use of an agonist of neurokinin B or neurokinin B in the reduction of blood volume in cases of hypotension.

10 In a twenty-first aspect of the invention there is provided a method of alleviating pre-eclampsia in a human subject by modifying the diet of the human subject to reduce the content of toxin generating substances therein.

15 In a twenty-second aspect of the invention there is provided a method of alleviating pre-eclampsia in a human subject including modifying the dietary pattern of the subject to reduce concentrations of potential toxins in the portal vein.

20 In a twenty-third aspect of the invention there is provided a dietary methodology for the alleviation of pre-eclampsia in a human subject in which the amount of toxin generating substances is reduced.

Figure 1 shows the polypeptide sequence of cloned human neurokinin B precursor, available under Accession No. aaf76980

25

Figure 2 shows the polypeptide sequence of the active neurokinin B peptide.

Figure 3 shows the polynucleotide sequence of placental cDNA of the human neurokinin B precursor, where ATG is the initiation codon; TAG is the stop codon, AATAAA is a polyadenylation signal; AAAAA is the polyA tail; and GGCACAGAGCTGCTCCACAGGCACC is the PCR primer based on Homo sapiens cDNA clone 138761 (Accession No. R63635) similar to the bovine

30

WO 01/36979

PCT/GB00/04315

10

clone, of Accession No. P08858 neurokinin B precursor used to amplify complete gene.

Figure 4 shows the genomic sequence of neurokinin B, including the 27928
5 base pair promoter region, the introns, and seven exons (underlined).

Figure 5 shows the results of semi-quantitative PCR for the complete human neurokinin B precursor using mRNA collected at weeks 9, 13 and term. Reverse transcription PCR was performed using mRNA collected at weeks 9,
10 13 and term (T) to amplify a 733 bp full length neurokinin B precursor cDNA. Primers for β -actin were used as the controls (257 bp). M1 denotes a 1kb DNA ladder, and M2 denotes a 100 bp DNA ladder.

Figure 6 shows HPLC results for oxidised and reduced neurokinin B in
15 human pregnancy plasma and human term placenta. Placental extracts revealed the peptide to be present in significant amounts (21 pg g^{-1} in early and 25 pg g^{-1} in term placenta) and its chromatographic behaviour was identical to synthetic NKB. Partial oxidation of placental NKB during extraction resulted in the production of three oxidised forms in which one or
20 both of the two-methionine residues were oxidised (a in plasma and b in placenta). The resulting methionine sulfoxides conferred reduced hydrophobicity, so that they eluted before the reduced form. This elution pattern matched that produced by the partial oxidation of synthetic NKB by hydrogen peroxide. Complete oxidation by hydrogen peroxide resulted in all
25 the NKB eluting in the position of the first peak. A similar elution pattern was also observed after extraction of NKB from term placenta samples (b).

Figure 7 shows the cardiovascular effect of neurokinin B in conscious rats. Changes in blood pressure and heart rate during infusion of saline or
30 incremental doses of NKB in conscious unrestrained female rats. NKB was infused at doses of 1.8 nmol h^{-1} (per kg) from time = 0, 18 nmol h^{-1} (per kg) from time = 16 h and 180 nmol h^{-1} (per kg) from time = 20 h. Values are mean

WO 01/36979

PCT/GB00/04315

11

± s.e mean. * denotes a significant difference from the original baseline and from the values at t = 20 h (Friedman's test).

5 Figure 8 shows an *in situ* hybridisation of for neurokinin B mRNA in the placenta of humans and rats. a, human at term (39 weeks) with human antisense probe b, human at term (39 weeks) with human sense probe c, rat 18 day placenta with rat antisense probe and d, high magnification showing giant cells of the rat placenta expressing neurokinin B. Magnification: a, 10x original size, b 10x, c 16x, d 40x.

10

The present invention is partly based upon the discovery that early and/or excessive release of neurokinin B into the maternal blood stream by the developing placenta can be a cause of pregnancy induced hypertension and pre-eclampsia. In particular, it has been postulated that those likely to suffer from pregnancy induced hypertension or pre-eclampsia have slightly elevated levels of neurokinin B in the maternal blood stream at approximately 10 to 12 weeks into pregnancy. Monitoring of neurokinin B early in pregnancy, for example at 10 to 12 weeks or before, is useful in predicting whether the individual is likely to suffer from pregnancy induced hypertension or pre-eclampsia later in pregnancy, and whether they are likely to suffer from pre-eclampsia related foetal complications such a foetal growth retardation, foetal hypoxia or miscarriage Measurement of neurokinin B levels after 10 to 12 weeks into pregnancy, for example at 18 weeks may enable the prediction to be confirmed and a diagnosis of pregnancy induced hypertension or pre-eclampsia or related foetal complications to be made. Further, it has been observed that the level of increase in neurokinin B levels after any initial prediction of hypertension or pre-eclampsia correlates with the future severity of the condition. In particular, it has been shown that a relationship exists between the degree of increase in neurokinin B and the future severity of the condition. These observations can be used in the prediction of the future severity of the condition. Also, other post-processing fragments of the human neurokinin B precursor may be involved in the

25
30

WO 01/36979

PCT/GB00/04315

12

development of those conditions. In addition, the production of neurokinin B and/or other fragments of human neurokinin B precursor may be associated with the development of hypertension in non-pregnant individuals.

- 5 In the present invention, foetal complications include any foetal condition which is related to pre-eclampsia. Specifically, foetal complications include foetal growth retardation, foetal hypoxia, pre-term labour, and in severe cases, miscarriage.
- 10 For the purpose of the present invention, neurokinin B precursor gene products include polynucleotide sequences encoding neurokinin B precursor or neurokinin B, and neurokinin B precursor polypeptides. Polynucleotide sequences include genomic or cDNA sequences, for example those of Figures 3 or 4, and RNA, preferably mRNA. Preferably, the neurokinin B
- 15 precursor polypeptides have the sequences shown in Figure 1. Fragments of neurokinin B precursor gene products are fragments which are derived from the precursor gene products and include the polynucleotide or polypeptide sequences encoding neurokinin B, fragments thereof, and other post-processing fragments of the precursor. Preferably the neurokinin B peptide
- 20 derived from the precursor has the sequence of Figure 2. Epitopic fragments or variants are those which comprise an amino acid sequence, typically of at least 4 residues, which constitutes a site to which the antibody can bind. A preferred epitopic fragment is the amino acid sequence DMHD of Figure 1.
- 25 Also included are variants of neurokinin B precursor gene products. Preferably, variants share at least 80%, at least 90%, at least 95%, at least 98% and most preferably at least 99 % sequence identity with the neurokinin B precursor gene products or fragments thereof, and preferably retain the same biological activity as the gene product or fragment.
- 30
- "% identity", as known in the art, is a measure of the relationship between two polypeptide sequences between two polypeptide sequences or two

WO 01/36979

PCT/GB00/04315

13

polynucleotide sequences, as determined by comparing their sequences. In general, the two sequences to be compared are aligned to give a maximum correlation between the sequences. The alignment of the two sequences is examined and the number of positions giving an exact amino acid or
5 nucleotide correspondence between the two sequences determined, divided by the total length of the alignment and multiplied by 100 to give a % identity figure. This % identity figure may be determined over the whole length of the sequences to be compared, which is particularly suitable for sequences of the same or very similar length and which are highly homologous, or over
10 shorter defined lengths, which is more suitable for sequences of unequal length or which have a lower level of homology.

Methods for comparing the identity of two or more sequences are well known in the art. Thus for instance, programs available in the Wisconsin Sequence
15 Analysis Package, version 9.1 (Devereux J. et al., Nucleic Acids Res. 12:387-395, 1984, available from Genetics Computer Group, Madison, Wisconsin, USA), for example the programs BESTFIT and GAP, may be used to determine the % identity between two polynucleotides and the % identity between two polypeptide sequences. BESTFIT uses the "local
20 homology" algorithm of Smith and Waterman (Advances in Applied Mathematics, 2:482-489, 1981) and finds the best single region of similarity between two sequences. BESTFIT is more suited to comparing two polynucleotide or two polypeptide sequences which are dissimilar in length, the program assuming that the shorter sequence represents a portion of the
25 longer. In comparison, GAP aligns two sequences finding a "maximum similarity" according to the algorithm of Needleman and Wunsch (J. Mol. Biol. 48:443-354, 1970). GAP is more suited to comparing sequences which are approximately the same length and an alignment is expected over the entire length. Preferably, the parameters "Gap Weight" and "Length Weight" used
30 in each program are 50 and 3 for polynucleotide sequences and 12 and 4 for polypeptide sequences, respectively. Preferably, % identities and similarities

WO 01/36979

PCT/CB00/04315

14

are determined when the two sequences being compared are optimally aligned.

Other programs for determining identity and/or similarity between sequences are also known in the art, for instance the BLAST family of programs (Altschul S.F. *et al* , J. Mol. Biol., 215:403-410, 1990, Altschul S.F. *et al* , Nucleic Acids Res., 25:289-3402, 1997, available from the National Center for Biotechnology Information (NCBI), Bethesda, Maryland, USA and accessible through the home page of the NCBI at www.ncbi.nlm.nih.gov) and FASTA (Pearson W.R. and Lipman D.J., Proc. Nat. Acad. Sci., USA, 85:2444-2448, 1988, available as part of the Wisconsin Sequence Analysis Package) Preferably, the BLOSUM62 amino acid substitution matrix (Henikoff S. and Henikoff J.G., Proc. Nat. Acad. Sci., USA, 89:10915-10919, 1992) is used in polypeptide sequence comparisons including where nucleotide sequences are first translated into amino acid sequences before comparison.

Preferably, the program BESTFIT is used to determine the % identity of a query polynucleotide or a polypeptide sequence with respect to a polynucleotide or a polypeptide sequence of the present invention, the query and the reference sequence being optimally aligned and the parameters of the program set at the default value.

The first, second, third and fourth aspects of the invention relate to methods of predicting or diagnosing pregnancy induced hypertension or pre-eclampsia or related foetal complications in a human subject. These methods include, for example, assessing the concentration in a biological sample of neurokinin B precursor gene products, or variants or fragments thereof. These methods preferably comprise comparing the results of an assessment of the concentration of human neurokinin B gene product (e.g. neurokinin B or its precursor) in a sample with expected values or with the values found in the subject at an earlier date

WO 01/36979

PCT/CB00/04315

15

Preferably these methods are carried out at an early stage of pregnancy, for example at 10-12 weeks for prediction, or 18 weeks for diagnosis.

- 5 These methods may include any means of measuring neurokinin B gene products available to those skilled in the art. Preferably, the methods use the kits of the invention. The methods of the invention comprise at least the step of determining the presence of neurokinin B mRNA, neurokinin B or its precursor, or variants or fragments thereof, in a biological sample; however,
- 10 additional steps may also be included. Such additional steps may include one or more of the following: collecting the biological sample; preparing the biological sample; measuring the concentration of target neurokinin B gene products such as polypeptide or polypeptides in the sample; preparing standard curves to predict expected concentrations of the target neurokinin B
- 15 gene products in non-pregnant individuals or in pregnant individuals at the same or different stages of pregnancy; comparing the results obtained from a particular biological sample with the appropriate expected values or the appropriate standard curve to determine the severity of the condition; or repeating some or all of the previous steps at a later date to determine if the
- 20 severity of the condition has changed.

- Suitable methods of detection based on kits will be clear to one skilled in the art and include radioimmunoassay (RIA), enzyme linked immunosorbant assay (ELISA), immunoradiometric assay (IRMA), antisense technology, or
- 25 radioreceptor assay (RRA). In the latter, for example the NK₃ receptor or other neurokinin B binding partner may be used in a detection system or biosensor system. Further detection methods may also include as well as radiometric methods, non-radioactive methods such as fluorescence and luminescence.

30

A preferred method is radioimmunoassay, which relies on the interaction of a small amount of radiolabeled peptide, eg neurokinin B, with a limiting amount

WO 01/36979

PCT/GB00/04315

16

of binding partner such as antibody (e.g. specific for NKB). The displacement of radiolabeled peptide by increasing doses of standard peptide is compared to that displaced by unknowns. This is normally monitored by separating binding partner bound label from free label usually by using a precipitation
5 step which brings down the binding partner followed by centrifugation, although there are adsorbents (e.g. charcoal) which can bind the free labeled fraction and can then be removed by centrifugation. IRMA can be one site or two site and uses an excess of specific binding partner such as antibody which in this case is radiolabeled. In the one site assay, separation is
10 effected by an excess of peptide linked to a solid phase which removes unreacted binding partner. In the two site method a second specific binding partner (usually linked to a solid phase) is used which is specific to a separate epitope on the peptide. Separation is easily effected by removal of the complex on the solid phase. RRA is similar to RIA in that a limiting
15 amount of receptor is substituted for the antibody. Often the receptor preparation will be in the form of a membrane preparation so that washing and separation of the bound label can be performed by e.g. centrifugation. The use of enzymes as the signalling moiety in immunometric assays is commonly achieved by cross linking an enzyme to the specific antibody or
20 the use of e.g. a pig anti mouse antibody cross-linked to an enzyme when a mouse monoclonal antibody is used in the initial reaction.

The above methods may also be used in estimating the likely future degree of pregnancy induced hypertension or pre-eclampsia or related foetal
25 complications. These methods preferably comprise comparing the results of an assessment of the concentration of human neurokinin B gene product (e.g. neurokinin B or its precursor) in a sample with expected values. It is believed that the tenth week of pregnancy, or later, for example after 18 weeks, may be particularly valuable times at which to assess the presence
30 (and concentration) of the human neurokinin B gene products.

WO 01/36979

PCT/GB00/04315

17

The methods of the invention are preferably carried out *in vitro*, on a sample removed from the body. Any biological sample may be used in the methods of the invention. Preferred biological samples include blood, saliva or urine

- 5 The invention also provides a method of preventing or treating pregnancy induced hypertension or pre-eclampsia or related foetal complications in a human subject by the administration of an agent which inhibits the biological effect of neurokinin B. Preferably, such methods are carried out using the kits of the invention. Agents which inhibit the biological effects of neurokinin
- 10 B include any agents that act, for example, by removing the neurokinin B from the plasma; by altering its structure to prevent it binding to receptors; by binding to the receptors directly to block the binding of neurokinin B thereto (but without themselves causing the effects at those receptors normally caused by neurokinin B), by exerting a counter effect to the neurokinin B at
- 15 the same or different receptors or by reducing or preventing gene expression or translation, for example by modulating activity of the neurokinin B gene promoter and/or by using antisense technology. Also included are agents which inhibit the production or processing of the precursor to prevent production of neurokinin B. Within this context, agents inhibiting the
- 20 biological effect of neurokinin B include agents inhibiting the biological effect of any variants or fragments of human neurokinin B or its precursor which are involved in the development of pregnancy induced hypertension or pre-eclampsia or related foetal complications. The principal site of action of human neurokinin B is the NK₃ receptor and therefore preferred agents
- 25 which inhibit the biological effects of neurokinin B for use in the invention include NK₃ receptor antagonists. However, at the high circulatory concentrations found in near term pregnancy, particularly in pregnancy induced hypertensive or pre-eclamptic subjects, neurokinin B may also have significant effects at other receptors (eg the NK₁ or NK₂ receptors) and
- 30 therefore the agents which inhibit the biological effects of neurokinin B for use in the present invention also include agents which prevent neurokinin B's

WO 01/36979

PCT/CB00/04315

18

effects at such other specific receptors, as well as broad spectrum neurokinin antagonists and combinations thereof.

Since 1991, a number of high-affinity nonpeptide antagonists have been reported. Snider R. M., et al., (Science, 251:435 (1991)), and Garret C., et al., (Proc. Natl. Acad. Sci., 88:10208 (1991)), described CP-96,345 and RP 67580, respectively, as antagonists at the NK₁ receptor, while Advenier C., et al., (Brit. J. Pharmacol., 105:78 (1992)), presented data on SR 48968 showing its high affinity and selectivity for NK₂ receptors. More recently
10 Macleod, et al., (J. Med. Chem., 36:2044 (1993)) have published on a novel series of tryptophan derivatives as NK₁ receptor antagonists. Recently, FK 888, a "dipeptide" with high affinity for the NK₁ receptor was described (Fujii J., et al., Neuropeptide, 22:24 (1992)).

15 Suitable NK₃ receptor antagonists for use in the present invention include all materials blocking or reducing the effect of neurokinin B at the NK₃ receptor, for example, those materials described in Gao and Peet (Current Medicinal Chemistry, 1999, 6, 375-388), Khavaga and Rogers (Int.J.Biochem Cell Biol. 1996, 28, 7, 721-738), US 5,942,523, US 5,846,973, US 5,491,140, US
20 5,328,927, US 5,360,820, US 5,344,830, US 5,331,089, US 4,742,156, US 4,665,157, EP 591,040A, WO 94/01402, WO 94/04494, WO 93/011609, Canadian Patent Application 2,154,116, EP 693,489 and Canadian Patent Application 2,151,116. Specific examples of suitable antagonists include the receptor selective ligand, SR 142801 (Edmonds-Ait, et al., Life Sciences,
25 56:27 (1995)), and the decapeptides of formula: A¹-D-Pro²-His³-D⁴-Phe⁵-D-Trp⁶-Val⁷-D-Trp⁸-Leu⁹-Nle¹⁰-NH₂ wherein A¹ and D⁴ are Asp or D-Asp amino acids.

Preferred agents for inhibiting the biological effects of neurokinin B include
30 those which modulate activity of the neurokinin B precursor gene promoter, thus altering the level of transcription of the neurokinin B precursor gene. Examples of such agents include competitive or non-competitive antagonists

WO 01/36979

PCT/GB00/04315

19

- of neurokinin precursor B gene promoter transcription factors, agents which inhibit the biological effect of neurokinin B precursor gene promoter transcription factors, agonists of neurokinin B precursor gene promoter inhibitors, and polynucleotide sequences which bind to, and inhibit, neurokinin B precursor gene promoter activity. Preferably, such polynucleotide will be sufficiently complimentary to whole or part of the promoter sequence such that they hybridise thereto and inhibit promoter activity, preferably *in vivo*. Examples of suitable polynucleotide sequences are those which have at least 80%, 85%, 90%, 95%, 97%, 98% and preferably 99% sequence identity with the compliment of whole or part of the promoter. Preferably the polynucleotide sequence will be complimentary to a regulatory region of the promoter, for example a transcription factor binding site.
- Where the agent is a polynucleotide sequence, it is preferably administered in the form of a vector. The vector may additionally comprise one or more regulatory sequences for activation of expression of the polynucleotide sequence, for example promoters including response elements, consensus sites, methylation sites, locus control regions, post-transcriptional modifications, splice variants, homeoboxes, inducible factors, DNA binding domains, enhancer sequences, initiation codons, and polyA sequences. Such agents may be administered by any suitable gene therapy technique, which will be known to persons skilled in the art.
- Administration of pharmaceutical compositions is accomplished by any effective route, e.g. orally or parenterally. Methods of parenteral delivery include topical, intra-arterial, subcutaneous, intramedullary, intravenous, or intranasal administration. Administration can also be effected by amniocentesis related techniques. Oral administration followed by subcutaneous injection would be the preferred routes of uptake; also long acting immobilisations would be used. Also, as the effects of placental NKB will be on peripheral receptors, effectively drugs devoid of side effects to the

WO 01/36979

PCT/CB00/04315

20

central nervous system should be preferably peptide-like in their distribution properties. In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically acceptable carriers comprising excipients and other compounds that facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of "REMINGTON'S PHARMACEUTICAL SCIENCES" (Maack Publishing Co, Easton PA).

10 Pharmaceutical compositions for oral administration can be formulated using pharmaceutically acceptable carriers well known in the art, in dosages suitable for oral administration. Such carriers enable the pharmaceutical compositions to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions, etc., suitable for ingestion by the patient.

15 Pharmaceutical preparations for oral use can be obtained through combination of active compounds with solid excipient, optionally grinding a resulting mixture, and processing the mixture of granules, after adding suitable additional compounds, if desired, to obtain tablets or dragee cores.

20 Suitable excipients are carbohydrate or protein fillers. These include, but are not limited to sugars, including lactose, sucrose, mannitol, or sorbitol, starch from corn, wheat, rice, potato, or other plants; cellulose such as methyl cellulose, hydroxypropylmethyl-cellulose, or sodium carboxymethylcellulose; and gums including arabic and tragacanth; as well as proteins, such as

25 gelatin and collagen. If desired, disintegrating or solubilising agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, alginic acid, or a salt thereof, such as sodium alginate.

Dragee cores are provided with suitable coatings such as concentrated sugar solutions, which may also contain gum arabic, talc, polyvinylpyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may

30

WO 01/56979

PCT/CB00/04315

21

be added to the tablets or dragee coatings for product identification or to characterise the quantity of active compound (i.e. dosage).

Pharmaceutical preparations, which can be used orally, include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a coating such as glycerol or sorbitol. Push-fit capsules can contain active ingredients mixed with filler or binders such as lactose or starches, lubricants such as talc or magnesium stearate, and, optionally, stabilisers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid paraffin, or liquid polyethylene glycol with or without stabilisers.

Pharmaceutical formulations for parenteral administration include aqueous solutions of active compounds. For injection, the pharmaceutical compositions of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hank's solution, Ringer's solution, or physiologically buffered saline. Aqueous injection suspensions may contain substances, which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Optionally, the suspension may also contain suitable stabilisers or agents, which increase the solubility of the compounds to allow for the preparation of highly concentrated solutions.

For topical or nasal administration, penetrants appropriate to the particular barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

30

The pharmaceutical compositions of the present invention may be manufactured in a manner similar to that known in the art (e.g. by means of

WO 01/36979

PCT/GB00/04315

22

conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping or lyophilising processes). The pharmaceutical compositions may also be modified to provide appropriate release characteristics, e.g. sustained release or targeted release, by convention means, e.g. coating.

The pharmaceutical composition may be provided as a salt and can be formed with many acids, including but not limited to hydrochloric, sulfuric, acetic, lactic, tartaric, malic, succinic, etc. Salts tend to be more soluble in aqueous or other protonic solvents than are the corresponding free base forms. In other cases, the preferred preparation may be a lyophilised powder in 1 mM-50 mM histidine, 0.1%-2% sucrose, 2%-7% mannitol at a pH range of 4.5 to 5.5, that is combined with buffer prior to use.

The agents for use in the invention (eg NK_3 receptor antagonists) can also be modified so that they are only delivered to selected target sites. For example, by adjusting their stability towards proteolytic digestion in the gut or ability not to pass the blood/brain barrier, or by producing composite molecules including a targeting component, e.g. an antibody selective for the target site.

After pharmaceutical compositions comprising a compound of the invention formulated in an acceptable carrier have been prepared, they can be placed in an appropriate container and labelled for treatment of an indicated condition. For administration of NK_3 receptor antagonists, such labelling would include amount, frequency and method of administration.

Pharmaceutical compositions suitable for use in the present invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. Thus, a therapeutically effective amount is an amount sufficient to ameliorate the symptoms of the disease being treated. The amount actually administered will be dependent upon the

WO 01/36979

PCT/GB00/04315

23

individual to which treatment is to be applied, and will preferably be an optimised amount such that the desired effect is achieved without significant side-effects. The determination of a therapeutically effective dose is well within the capability of those skilled in the art. Of course, the skilled person
5 will realise that divided and partial doses are also within the scope of the invention.

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays or in any appropriate animal model (eg
10 primates for pre-eclampsia, rats and guinea pigs for hypertension and other small laboratory animals for use with induced hypertension and induced pre-eclampsia). These assays should take into account receptor activity as well as downstream processing activity. The animal model is also used to achieve a desirable concentration range and route of administration. Such
15 information can then be used to determine useful doses and routes for administration in humans

A therapeutically effective amount refers to that amount of agent, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity of
20 such compounds can be determined by standard pharmaceutical procedures, in cell cultures or experimental animals (e.g. ED_{50} , the dose therapeutically effective in 50% of the population; and LD_{50} , the dose lethal to 50% of the population). The dose ratio between therapeutic and toxic effects is the therapeutic index, and it can be expressed as the ration ED_{50}/LD_{50} .
25 Pharmaceutical compositions, which exhibit large therapeutic indices, are preferred. The data obtained from cell culture assays and animal studies is used in formulating a range of dosage for human use. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED_{50} with little or no toxicity. The dosage varies within this range
30 depending upon the dosage form employed, sensitivity of the patient, and the route of administration.

WO 01/36979

PCT/GB00/04315

24

The exact dosage is chosen by the individual physician in view of the patient to be treated. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Additional factors, which may be taken into account, include the severity of the disease state. Long acting pharmaceutical compositions might be administered every 3 to 4 days, every week, or once every two weeks depending on half-life and clearance rate of the particular formulation. Guidance as to particular dosages and methods of delivery is provided in the literature (see, US Patent No's 4,657,760; 5,206,344 and 5,225,212 herein incorporated by reference).

10

The agents which inhibit the biological effect of neurokinin B for use in the methods of the invention of preventing or treating pre-eclampsia; or of preparing medicaments for preventing or treating pre-eclampsia; are preferably formulated such that use of the agent is effective in, but not restricted to, the post prandial phase. The agents may for example be selected to be effective over a 24 hour period rather than exclusively in the post-prandial phase. The post-prandial phase is a particularly important time as it is believed that pre-eclampsia is associated with the build-up of toxins in the maternal blood supply due to the failure of the blood to pass through the liver (which normally removes the toxins) because of high pressure in the portal vein. Thus, transient relief of hypertension following meals will allow the blood to pass through the liver at the time when the highest concentration of toxins will be present and will therefore provide a large reduction in the risk of pre-eclampsia whilst producing only a short decrease in the effect caused by the placentally produced neurokinin B. This time limited effect may be achieved by selecting agents with short durations of activity and using appropriate formulations and dosage schedules.

Preferably, methods of prevention or treatment of the conditions addressed herein will begin as soon as possible after the initial prediction or diagnosis is made, for example after 10 weeks into pregnancy. The decision regarding initiation of a course of treatment will of course be the decision of a physician,

30

WO 01/36979

PCT/GB00/04315

25

and may therefore begin earlier or later. Typically, the course will be given throughout pregnancy or until symptoms subside. This may continue until up to eight weeks after birth. In individuals who have been determined as being at risk of developing foetal conditions such as growth retardation or hypoxia, or pre-eclampsia, (by consideration of other factors such as previous miscarriages or complications in pregnancy) the course may be initiated as soon as pregnancy is confirmed, and may continue until term.

10 In a further aspect of the invention there is provided the use of a human neurokinin B precursor gene product or a variant or fragment thereof in the manufacture of a diagnostic for use in the prediction or diagnosis of pregnancy included hypertension or pre-eclampsia or related foetal complications. Preferably, the gene product used is neurokinin B, or a variant or fragment thereof, for example in the production of a diagnostic comprising a binding partner specific for neurokinin B. Preferably, the variants or fragments are epitopic. It is envisaged that other gene products could also be used, for example regulatory sequences of the neurokinin B precursor genomic sequence, or neurokinin B precursor mRNA in the production of antisense sequences

20 The polypeptides used include human neurokinin B or its precursor, or variants or fragments thereof. Preferably, the polypeptides comprise the sequence of Figure 1 or Figure 2 respectively. Preferably, the fragments or variants are epitopic, as defined above.

25 These polypeptides may be produced in isolated, substantially pure form or as recombinant polypeptides. Method for doing so will be clear to one skilled in the art. These will include, for example, recombinant techniques or extraction, gel separation or more commonly, for peptides the size of neurokinin B, chemical synthesis, eg liquid and solid phase peptide.

30

WO 01/36979

PCT/GB00/04315

26

In a further aspect of the invention there is provided the use of an agent which inhibits the biological effect of neurokinin B in the manufacture of a medicament for the prevention or treatment of pregnancy induced hypertension or pre-eclampsia or related foetal complications. Preferably, the agents are those defined above.

In a further aspect of the present invention there are provided kits for the predicting the onset of, diagnosing, or estimating the future severity of pregnancy induced hypertension or pre-eclampsia or related foetal complications. The kits of the invention comprise a means for detecting the production of human neurokinin B gene products such as polynucleotides or polypeptides encoding neurokinin B or its precursor, or fragments or variants thereof, by the subject. Thus the kits will commonly comprise one or more of: a binding partner to neurokinin B or its precursor; neurokinin B polypeptide or variants or fragments thereof; and/or polynucleotide sequences which hybridise to a sequence encoding neurokinin B or a variant or fragment thereof.

By binding partner is meant any substance capable of detecting (and binding to) the target, eg an antibody. Preferred binding partners for use in the kits of the invention are antibodies which are specific for neurokinin B precursor, or epitopic fragments or epitopic variants thereof. Preferred are antibodies to neurokinin B and antibodies to the human neurokinin B precursor. Most preferred are antibodies which are specific for neurokinin B, but antibodies specific to any other breakdown products of the neurokinin B precursor which remain in the body for a measurable time may also be used. These antibodies are capable of binding fragments of the human neurokinin B precursor to identify the production of the precursor by the human body. The antibodies of the invention may be, for example, polyclonal, monoclonal, chimeric or humanised antibodies or fragments thereof. Binding partners which cross react with related peptides such as Substance P or NKA, for

WO 01/36979

PCT/GB00/04315

27

example, may be useful as a medicament or in diagnosis, as they share a common sequence (FVGLM-NH₂) with neurokinin B

Methods of producing such antibodies will be apparent to one skilled in the art. For example, in the case of polyclonal antibodies, by standard methods of animal immunisation or, for monoclonal antibodies, by the well-known methods of Köhler and Milstein, or by use of the methods discussed in US 5,844,080. Chimeric antibodies can be made by genetic engineering techniques, and are antibodies in which the constant region is human in origin, but the variable regions are derived from, for example, a mouse antibody. The advantage of chimeric antibodies is to reduce immunogenicity. Humanised antibodies take this principle even further, in that only the complementarity determining regions and a minimum number of further amino acids in the variable regions are derived from an animal such as a mouse. The rest of the antibody structure is human in sequence, and is recognised by the human immune system as human (see, for example, Queen et al, PNAS, USA 86 (December 1989), 10029-10033).

Polynucleotides of the kits of the invention are preferably those which hybridise to a sequence encoding neurokinin B or its precursor, or a variant or fragment thereof, or complements thereof, under stringent conditions. Preferred are polynucleotide sequences which hybridise to the nucleotide sequence of Figure 3 or Figure 4, or their complements, under stringent hybridisation conditions. Stringent conditions are, for example, 6x SSC at 65°C. Preferably, such polynucleotide sequences have at least 85%, and least 90%, at least 95%, preferably at least 98% and most preferably at least 99% sequence identity with the complement of the reference sequence. Such polynucleotide sequences are preferably at least 10 nucleotides in length, and will be useful in detecting expression of neurokinin B or its precursor. Such polynucleotides are useful in antisense technology or diagnostic PCR.

WO 01/36979

PCT/GB00/04315

28

Means of producing the polynucleotides of the invention will be clear to those skilled in the art, for example, they may be produced synthetically or by probing an appropriate cDNA or genomic library (particularly a placental cDNA library).

5

The kits of the invention may also comprise instructions for the performance of an assay for predicting or diagnosing the levels of neurokinin B in a biological sample (this may either be by direct measurement of neurokinin B or by measuring the concentration of human neurokinin B precursor, or a fragment thereof, and using this value to predict the amount of neurokinin B present). The components of the commercial neurokinin B radioimmunoassay kit RIK 7357 by Peninsula Laboratories, Belmont, CA, USA can be used in the present invention. The kits of the invention preferably also comprise a key, showing the correlation between the levels of neurokinin B gene product in the biological sample and diagnosis of pregnancy induced hypertension or pre-eclampsia or related foetal complications, and/or the likely future onset and/or severity of these conditions.

Also provided are kits for the prevention or treatment of pregnancy induced hypertension or pre-eclampsia or related foetal complications, comprising means for inhibiting the biological effect of neurokinin B or its precursor in a subject. Preferably, such means include those agents defined above. In particular, the antibodies or polynucleotide sequences as described above may also be useful in these kits for inhibiting the biological effect of neurokinin B or its precursor. The kits preferably also contain instructions for use of the kit to prevent or treat pregnancy induced hypertension or pre-eclampsia or related foetal complications and/or a key showing the correlation between the amount of agent used and the likely effect on the condition.

WO 01/36979

PCT/GB00/04315

29

Pre-eclampsia may also be alleviated by modifying the diet of a human subject to reduce the content of toxins (e.g. alkaloids) and toxin generating substances therein. Toxin generating substances include proteins which are digested in, and absorbed from, the gut as amino acids most of which are toxic if they circulate in blood in too high concentrations. Normally any amino acids in excess of daily requirement are immediately deaminated by the liver and metabolised. Increasing the proportion of carbohydrates in the diet may also be of particular benefit. The dietary pattern of the subject may also be modified to prevent peak concentrations of potential toxins appearing in the portal vein, for example by substantially reducing the size of individual meals (and increasing the frequency of small meals).

Agonists of neurokinin B may also be used as pharmaceutical agents where an increase in blood pressure or decrease in blood volume is considered to be beneficial. Suitable agonists include any acting to supplement or mimic the effect of neurokinin B at the NK₃ receptor (or at any other receptor), for example senktide or [MePhe⁷] NKB.

The present invention also provides means of screening potential effective agents (eg NK₃ receptor antagonists and agonists) by testing their ability to block (or enhance) the hypertensive effect of neurokinin B in an appropriate model. Once suitable agents have been identified, they may then further be tested to determine their potential in preventing or treating hypertension; pregnancy induced hypertension or pre-eclampsia, and used accordingly. All agents identified by such a process (other than presently known materials) are included in the present invention. Screening methods include large array techniques such as the Vilsips™ technology of Affymetrix Inc; see, eg, EPB No 0476014.

30

Transfected cells lines containing the cloned NK₃ (or NK₁ or NK₂) receptor could be used in receptor binding and cell signalling pathway studies in a

WO 01/36979

PCT/G800/04315

30

way clear to one skilled in the art. Essentially, either cells lines expressing endogenously high levels of neurokinin receptors or cell lines transfected with cloned cDNA constructs of the neurokinin receptor may be used to produce membrane preparations. Membrane preparations, of purified receptors in solution or after reconstitution into phospholipid membranes, may then be used to assess receptor binding with labelled agonists and/or antagonists of neurokinin B. The effects of the action of the agonists and antagonists can be assessed using standard cell signalling assays. These will be typical of those routinely performed when using G-protein coupled receptors systems in a way clear to one skilled in the art (including such assays as receptor binding, cyclic AMP determination, protein kinase C, inositol triphosphate concentrations etc.). These studies could also be performed in animal models including the guinea pig and rat chronically infused with agonist to determine the long and short-term effects of neurokinin B, neurokinin B agonists and neurokinin B antagonists. Effects such as changes in heart rate, blood pressure, blood volume and weight of internal organs (e.g. uterus, placenta) may be measured.

EXAMPLES

20

Example 1

Production of human neurokinin B precursor cDNA

The cloning of placental cDNA, using the following methods, was used to identify the human neurokinin B precursor having the polypeptide sequence shown in Figure 1. The peptide sequence of neurokinin B in the precursor is underlined (the C-terminal G residue ends up as the amide on the C-terminal M in the final processed peptide of Figure 2). The cloned placental cDNA of the human neurokinin B precursor is shown in Figure 3 and has (underlined) the ATG initiation codon at 26-28, the TAG stop codon at 389-391, the AATAAA polyadenylation signal at 659-663 and the polyA tail starting at 680.

WO 01/36979

PCT/GB00/04315

31

- Human placental tissue was obtained from pregnancy terminations at weeks 9 and 13 of gestation and term. Samples were collected in compliance with and approval from the Local Research Ethics Committee. RNA was extracted essentially as described by Chomczynski, P. and Sacchi, N. (1987)
- 5 Analytical Biochemistry, 162, 156-159.

- The full-length preproneurokinin B precursor was amplified using RT-PCR from total human term placental RNA. This was done using the SMART RACE cDNA amplification method (Chenchik, A. et al (1998)). In RT-PCR
- 10 Methods for Gene Cloning and Analysis. Eds. Siebert, P. and Lamck, J. (BioTechniques Books, MA), 305-319). Essentially, after total RNA extraction, reverse transcription was performed using a cDNA synthesis primer (5'AAGCAGTGGTAACAACGCAGAGTAC(T)₃₀N₁N3') which contained a 3' anchor sequence. 3' race was performed using a 5' gene specific primer
- 15 (5'GGCACAGAGCTGCTCCACAGGCACCAT 3') derived from the Homo sapiens cDNA clone 138761 similar to bovine P08858 neurokinin B precursor. The resulting PCR fragment was gel purified following gel electrophoresis and cloned into the expression vector pGEM-T Easy. The resulting clones were sequenced and compared to submitted sequences in
- 20 the GenBank database using the BLAST program (Altschul, S.F , et al (1990) J.Mol Biol. 215 403-410).

Example 2

Semi-Quantitative PCR to measure NKB in placenta

- 25 Semi-quantitative PCR as described below was used to measure the mRNA expression of neurokinin B in placenta collected at 9 weeks, 13 weeks and at term. This showed differences in a degree of expression between the first trimester and term placenta. Expression levels were up by five times at term, as shown in Figure 5.

30

SMART RACE placental cDNA was amplified using a 5' gene specific primer (5'GGCACAGAGCTGCTCCACAGGCACCAT 3') derived from the Homo

WO 01/36979

PCT/GB00/04315

32

sapiens cDNA clone 138761 similar to bovine P08858 neurokinin B precursor and a 3' SMART anchor sequence primer. A specified primer pair for β -actin was used for normalisation. PCRs were performed using twenty-one cycles of 95°C for 30 sec and 68°C for 2 min. The primers were chosen deliberately to have high annealing temperatures so that the PCR reactions could be performed two step to reduce the possibility of non-specific products being formed. The number of cycles required to obtain a reproducible exponential amplification of the β -actin RT-PCR product was determined by terminating control reactions at 15, 18, 21, 24 and 30 cycles respectively. These experiments were used to check the accuracy, efficiency and amount of total RNA needed to obtain a semi-quantitative amplification in order to optimise the levels of β -actin PCR product produced. The PCR products were visualised by UV illumination following electrophoresis. (A 1kb DNA ladder (M1) and 100bp DNA ladder (M2) are shown in Figure 5 also)

Example 3

Neurokinin B extraction from placental tissue and plasma

Testing of placental extracts using the techniques set out below revealed neurokinin B to be present in significant amounts and its chromatographic properties in HPLC were identical to synthetic neurokinin B. It also displayed the same degree of loss of hydrophobicity (on HPLC) after oxidising its methionine residues. Oxidisation was found to give three peaks of double oxidised (1), single oxidised (2) and non-oxidised forms (3), see Figure 6. Figure 6(a) shows oxidised and reduced neurokinin B separated by RPHPLC from human pregnancy plasma and Figure 6(b) shows separation of condensed and reduced neurokinin by RPHPLC extracted from human term placenta.

Extraction of neurokinin B from placenta

Whole placentae were weighed and washed immediately after delivery with 150 mM sodium chloride solution containing 10 mM EDTA at pH 7.5. A

WO 01/36979

PCT/GB00/04315

33

tissue sample not exceeding 100g was excised and homogenised in 100 ml saline/EDTA solution using a blender with a glass vessel. Protease inhibitors, phenylmethylsulphonylfluoride, N-ethylmaleimide, and pepstatin were added from a stock solution in methanol. After 20 seconds 800 ml of methanol were added and blending was continued for a further minute. The mixture was decanted into 200 ml polypropylene centrifuge tubes and subjected to centrifugation at 4°C and 3000 X g for 30 minutes. The supernatant was separated and stored overnight at 4°C resulting in further precipitation that was removed by centrifugation. The volume of each extract was reduced to less than one eighth of the initial volume and then diluted by addition of three volumes of water containing 0.1% trifluoroacetic acid (TFA). Any trace of suspended matter was removed by a final centrifugation step. The volume of extract was recorded and an amount corresponding to 20g of placenta reserved for solid phase extraction using Sep-Pak C18 3CC cartridges (Waters Chromatography Division, Millipore Corporation, Milford, MA, U.S.A.). Cartridges were primed prior to use by perfusion with 2 ml of the following solutions; 1) water containing 0.1% TFA and 0.1% Polypep gelatine hydrolysate (Sigma-Aldrich, Poole, UK), 2) water containing 0.1% TFA, 3) water containing 80% v/v acetonitrile and 4) water containing 0.1% TFA. Each extract was passed through a prepared cartridge, which was then washed with 2 ml 0.1% TFA in water, 2 ml 0.1% TFA in water containing acetonitrile 10% and 20% TFA. The column was eluted with 2 ml of 30%, 40% and 50% acetonitrile in water containing 0.1% TFA. Eluted fractions were reduced to dryness under vacuum after adding 1 mg of mannitol and 100 µg Polypep. Smaller placentae obtained from abortions were treated as above but dissociated in a glass homogeniser retaining the same proportions of buffer and methanol to placental weight.

Extraction of neurokinin B from plasma

Neurokinin B standards were prepared in pooled plasma from the blood of five young males taken into EDTA. The standards contained 1280, 640, 320,

WO 01/36979

PCT/GB00/04313

34

160 and 80 pg/ml neurokinin B. Each 2ml of sample of plasma standard was acidified by addition of 220 μ l 1M HCl containing 0.21M glycine. They were then diluted to 10 ml with 0.9% saline and subjected to centrifugation at 3000 X g for 20 minutes to ensure complete clarity. Sep-Pak C18 1CC cartridges were primed as described above for Sep-Pak C18 3CC cartridges. After loading, cartridges were washed with 1 ml 0.1 M HCl containing 0.02M glycine followed by 1 ml 0.1% TFA in water. Further washes with 1ml 0.1% TFA in water containing 10 and 20% acetonitrile were followed by elution with 1 ml 0.1% TFA in a mixture of 50% water and acetonitrile. Eluted fractions were reduced to dryness under vacuum after adding 1 mg of mannitol and 100 μ g Polypep. The acidification step ensured that we were extracting already processed mature peptide as it is possible that inactive circulating precursor could be cleaved by endogenous plasma proteases to produce immunoreactive peptides unless precautions are taken.

15

Example 4

Measurement of NKB in placental tissues and plasma

Placental and plasma extracts were reconstituted in 500 μ l of buffer supplied as part of a commercial neurokinin B radioimmunoassay kit RIK 7357 by Peninsula Laboratories, Belmont, CA, USA to which had added 0.2% Igepal CA-630 non-ionic detergent (Sigma). Sub-samples of 25 μ l were taken from extracted and non-extracted standards and mixed with 75 μ l of the above buffer. Standards were prepared in buffer containing Igepal, but to which had been added 200 μ g/ml Polypep. Anti-neurokinin antibody solution (100 μ l) was added to all assay tubes except blanks and the assay was conducted as described in the "General Protocol for Radioimmunoassay Kit" instructions. Assays were performed in duplicate and results were corrected with reference to extracted standards.

30

WO 01/36979

PCT/GB00/04315

35

The plasma and placental levels of neurokinin B in various human volunteers and rats were measured by the above methods. The results of the plasma samples are summarised in Table 1. Placental samples were collected from weeks 7 to 15 of pregnancy, and all seven were shown to contain equivalent significant amounts of neurokinin B; however concentrations of plasma NKB detected at term were in the 100 picomolar range that would be expected to have effects on the maternal cardiovascular system. Plasma samples taken from non-pregnant volunteers all had low levels of the peptide, as did the majority of plasma samples taken from individuals who had been admitted for elective abortions at weeks 7 to 15. Four samples from this latter group had concentrations equivalent to those found at term. This suggests that the placenta from this individual may have started to secrete supra-physiological concentrations of neurokinin B early in pregnancy. Samples of patients in late pregnancy suffering from hypertension and pre-eclampsia all had concentrations in the nanomolar range suggesting that raised neurokinin B may be responsible for their symptoms.

WO 01/36979

PCT/GB00/04313

36

Table 1

Week of Pregnancy	Nmol/l NKB in normotensive pregnancies
6	0
9	0
9	0.97
10	0.535
13	0
13	0
13	0.083
13	0.511
14	0
14	0
14	0.511
17	0.182
17	0.182
18	0
23	0.12
24	0
25	0.17
27	0
28	0
28	0.033
31	0
31	0.031
32	0
33	0
37	0
38	0.07
39	0.138
40	0.05
40	0.2
41	0.118

WO 01/36979

PCT/GB00/04315

37

Table 2

Week of pregnancy	Nmol/l NKB in pre-eclamptic pregnancies
30	3.964
34	6.156
36	3.796
37	2.141
38	2.752
39	2.004
39	6.288
39	0.98

5 Table 3

Patient number	Nmol/l NKB in normotensive pregnancies at term
1	0
2	0
3	0
4	0
5	0
6	0
7	0
8	0.084
9	0.118
10	0.143
11	0.22
12	0.226
13	0.228
14	0.398
15	0.521
16	1.317

WO 01/36979

PCT/GB00/04315

38

CLAIMS:

1. A kit for the prediction or diagnosis of pregnancy induced
5 hypertension, pre-eclampsia or related foetal complications comprising a
binding partner, eg an antibody, to neurokinin B precursor gene product or
variant or fragment thereof.
2. A kit according to claim 1 further comprising instructions for the
10 performance of an assay for predicting the levels of neurokinin B in a
biological sample and correlating the assay results with the likely future
development of pregnancy induced hypertension or pre-eclampsia or related
foetal complications respectively
- 15 3 A kit for use in estimating the likely future degree of pregnancy
induced hypertension or pre-eclampsia or related foetal complications,
comprising a binding partner, eg an antibody, to neurokinin B precursor gene
product or variant or fragment thereof, together with instructions for the
20 performance of an assay for predicting the levels of neurokinin B in a
biological sample and correlating the assay results with the predicted future
severity of pregnancy induced hypertension or pre-eclampsia or related foetal
complications, respectively
- 4 A kit as claimed in any one of claims 1 to 3 wherein the binding partner
25 is an antibody specific for neurokinin B precursor, or neurokinin B or an
epitopic fragment or epitopic variant thereof.
5. A kit according to any one of claims 1 to 4 wherein the binding partner
is an antibody specific for the human neurokinin B precursor having the
30 sequence of figure 1 or an epitopic variant or epitopic fragment thereof.

WO 01/36979

PCT/G800/04315

39

6. A kit as claimed in any one of claims 1 to 5 which is a radioimmunoassay kit, an enzyme linked immunosorbant assay kit, an immunoradiometric assay kit or a radioreceptor assay kit.
- 5 7. A method of preventing or treating pregnancy induced hypertension or pre-eclampsia or related foetal complications in a human subject by the administration of an agent which inhibits the biological effect of neurokinin B
8. The method as claimed in claim 7 wherein the agent which inhibits the biological effect of neurokinin B is an NK₁, NK₂ or NK₃ antagonist.
- 10 9. The method as claimed in claim 8 wherein the NK₃ antagonist is a decapeptide with the following formula - A¹ -D-Pro² -His³ -D⁴ -Phe⁵ -D-Trp⁶ -Val⁷ -D-Trp⁸ -Leu⁹ -Nle¹⁰ -NH₂ wherein A¹ and D⁴ are Asp or D-Asp amino acids or SR 142801.
- 15 10. The method as claimed in claim 7 wherein the agent which inhibits the biological effect of neurokinin B is one which modulates the activity of the neurokinin B precursor gene promoter.
- 20 11. The method as claimed in any one of claims 7 to 10 wherein the agent is selected and administered such that it effective over a 24 hour period.
12. Use of human neurokinin B precursor gene product or variant or a fragment thereof in the manufacture of a diagnostic for use in the prediction or diagnosis of pregnancy-induced hypertension or pre-eclampsia or related foetal complications.
- 25 13. Use of a human neurokinin B precursor gene product or variant or fragment thereof according to claim 12, wherein the gene product is human neurokinin B precursor or human neurokinin B, or an epitopic variant or epitopic fragment thereof.
- 30

WO 01/36979

PCT/GB00/04315

40

- 14 Use of neurokinin B in the manufacture of a diagnostic for use in the prediction or diagnosis of pregnancy induced hypertension or the diagnosis of pre-eclampsia or related foetal complications, according to claims 12 or
- 5 13.
- 15 Use of an agent which inhibits the biological effect of neurokinin B in the manufacture of a medicament for the prevention or treatment of pregnancy induced hypertension or pre-eclampsia or related foetal
- 10 complications.
- 16 The use as claimed in claim 15 wherein the agent which inhibits the biological effect of neurokinin B is an NK₁, NK₂ or NK₃ antagonist.
- 15 17 The use as claimed in claim 15 wherein the NK₃ antagonist is SR 142801, or the decapeptides with the following formula : A¹ -D-Pro² -His³ -D⁴ -Phe⁵ -D-Trp⁶ -Val⁷ -D-Trp⁸ -Leu⁹ -Nle¹⁰ -NH₂ wherein A¹ and D⁴ are Asp or D-Asp amino acids
- 20 18 The use as claimed in claim 15 wherein the agent which inhibits the biological effect of neurokinin B is one which modulates activity of the neurokinin B gene promoter.
- 19 The use as claimed in any one of claims 15 to 18 wherein the
- 25 medicament is formulated such that the agent is effective over a 24 hour period.
- 20 A method of predicting or diagnosing pregnancy induced hypertension or pre-eclampsia or related foetal complications at an early stage in a human subject by assessing the concentration in a biological sample, e.g. blood, of human neurokinin B precursor gene product or a variant or a fragment thereof.
- 30

WO 01/36979

PCT/GB00/04315

41

21. A method of predicting or diagnosing pregnancy induced hypertension at an early stage in a human subject or of predicting pre-eclampsia or related foetal complications at an early stage in a human
5 subject by assessing the concentration in a biological sample, e.g. blood, of neurokinin B or its precursor.

22. A method according to claim 21 wherein neurokinin B and its precursor have the sequences of figures 1 and 2 respectively.

10

23 The method as claimed in claims 20 to 22 comprising the use of a kit as defined in any one of claims 1 or 2.

24. A method of estimating the likely future degree of pregnancy induced hypertension or pre-eclampsia or related foetal complications in a human
15 subject by assessing the concentration in a biological sample, eg blood, of human neurokinin B precursor gene product or a variant or a fragment thereof, and correlating the result with the predicted future severity of pregnancy induced hypertension ore pre-eclampsia or related foetal
20 complications.

25. A method according to claim 24 comprising assessing the concentration in a biological sample, e.g. blood, of nuerokinin B.

25 26. The method as claimed in any one of claims 24 or 25 comprising the use of a kit as defined in any one of claims 3 to 5.

27. The method as claimed in claim 26 wherein the kit comprises an antibody specific for neurokinin B.

30

WO 01/36979

PCT/CB00/04315

42

28. The method as claimed in claim 26 or claim 27 wherein the kit is a radioimmunoassay kit, an enzyme linked immunosorbant assay kit, an immunoradiometric assay kit or a radioreceptor assay kit.
- 5 29. The use of neurokinin B or an agonist thereof in the reduction of blood volume in cases of hypotension.
30. The use of neurokinin B or an agonist thereof in the preparation of a medicament for the reduction of blood volume in cases of hypotension
- 10 31. A method of alleviating pre-eclampsia in a human subject by modifying the diet of the human subject to reduce the content of toxin generating substances therein.
- 15 32. A method of alleviating pre-eclampsia in a human subject including modifying the dietary pattern of the subject to reduce concentrations of potential toxins in the portal vein.
- 20 33. A dietary methodology for the alleviation of pre-eclampsia in a human subject in which the amount of toxin generating substances is reduced.